Abstract—Multilayered valve-based continuous-flow microfluidic biochips are a rapidly developing platform for delicate bio-applications. Due to the high complexity of the biochip structure and the application protocols, there is an increasing demand for design automation approaches. Current research has enabled automated generation of biochip physical designs, operation scheduling, and binding protocols, which has demonstrated the potential for better resource utilization and execution time reduction. However, the state-of-the-art high-level synthesis methods are on operation- and device-level. They assume fluid transportation paths to be always available but overlook the physical layout of the control and flow channels. This mismatch leads to a gap in the complete synthesis flow, and can result in performance drop, waste of resources due to redundancy or even infeasible designs. This work proposes to bridge this gap with a simulation-based approach, which takes a biochip design and a high-level protocol as inputs, and synthesizes channel-level pressurization protocols to support dynamic construction of valid fluid transportation paths. Experimental results show that the proposed method can efficiently validate the dynamic construction of flow paths for feasible designs and protocols, detect redundant resource usage, and locate the conflicts for infeasible designs and protocols. It opens up a new direction to improve the performance and the feasibility of customized biochip synthesis.

I. INTRODUCTION

Multilayered continuous-flow microfluidic biochips [1] are a promising lab-on-a-chip platform for high-throughput biological applications. Soft lithography technique is applied to bond multiple patterned layers of elastomer, each of which consists of dedicated channels that allow gas or fluids to pass through. The multilayered structure enables the construction of active valves. A valve is a composite functional unit consisting of channel segments from different layers and a flexible membrane at the layer interface. A two-layer polydimethylsiloxane (PDMS) push-up valve [2] is shown in Figure 1, where a flow layer is on top of a control layer. When gas or oil infuses the bottom control channel, the channel will be pressurized and thus push the membrane upwards. Since the flow channel segment has a rounded profile that perfectly fits the expanded membrane, the channel will be sealed and the fluid movement in this channel will be blocked [3].

With channels and valves, bioengineers have been able to build delicate microfluidic systems on coin-sized chips. But as the complexity of the application protocols and the integration scale of the biochips increase, manually designing the chips becomes more and more time-consuming and error-prone, which results in a strong demand for automated software synthesis tools. Many design automation researchers put their efforts into analyzing and solving this design problem. The ambition is to construct a completely automated synthesis flow, which can transform a high-level abstraction of a given application into a feasible and optimized chip design with an explicit protocol for executing the application.

With a decade of effort, researchers have achieved significant progress covering both high-level synthesis and physical design. Specifically, up to 2019, there have been: high-level modeling methods that optimize the resource utilization based on given application protocols [4] [5], place-and-route tools that synthesize manufacturing-ready physical designs targeting applications of different scales [6] [7] [8], scheduling and fluid routing approaches that map operations to given physical topologies [9] [10], and other frameworks focusing on specific optimization criteria such as fluid storage [11] [12] and control pin reduction [13] [14], etc.

Though the proposed approaches gradually piece together the complete automatic synthesis flow, there are still some missing links between the designs and the protocols.

First of all, the state-of-the-art high-level synthesis methods are on operation- and device-level only. They assume fluid transportation paths to be always available but overlook the interaction between the control and flow channels. Specifically, flow path construction is a dynamic process in multilayered biochips. To transport fluids from one location to another, it is not sufficient to ensure the static physical connection of flow channels, but also necessary to pressurize certain control channels to temporally block the fluid movement in unwanted directions. Though current approaches can synthesize the operation scheduling and device binding results, they cannot produce the explicit control channel pressurization sequences to validate the dynamic construction of flow paths. Flow path validation is indispensable because of two reasons:

- Reason 1: flow path construction must not only consider the local channel structure, but should consider the whole chip. As pointed out in [10], to move fluids from location a to location b, three sub-paths need to be considered: a path from a to b, a path from an inlet to a, and a path from b to an outlet. Besides, as mentioned earlier, it is insufficient to ensure the physical connection of the corresponding flow channels, but also necessary to guarantee that all valves along the paths are properly pressurized and depressurized. Manually deriving
which intends to form a flow path from $i$ to $v_2$. (b) A fabricated chip [7] where valves A and B are connected to share the pressure.

Figure 2: (a) A manually derived valve actuation protocol from [13], which intends to transport fluids from an inlet $i$ to the bottom half-ring between valves $v_1$ and $v_2$, but mistakenly sets valves right to the dash-line to 'don’t care' status, indicating that these valves are irrelevant and can be either pressurized or not. However, if $v_3$ and $v_4$ are both pressurized and thus ‘closed’, there will be no sub-path from $v_2$ to an outlet. Since the initial status of a channel is not a vacuum but filled with air, it is hardly possible\(^1\) to form the expected pattern of fluid flow.

- Reason 2: different sub-paths may conflict with one another due to pressure-sharing valves. In multilayered biochips, valves are connected to external pressure sources via control pins, which are punch holes on the chip that occupy a large area.\(^2\)

To reduce the control pin usage, it is a common approach to connect multiple valves sequentially with the same control channel to share the same control pin. In this case, pressurizing one control channel will lead to the pressurization of all valves along the channel, which may result in unexpected blockage of flow paths. For example, Figure 2(b) shows the photo of a biochip [7] consisting of two ring-shaped mixers. The blue dash line demonstrates a path for fluid transportation from an inlet to the upper mixer. To prevent the bottom mixer from contamination, an intuitive approach is to pressurize valve A. However, since valve A and valve B are sequentially connected by a green control channel, pressurizing valve A leads to the pressurization of valve B, which results in the unwanted blockage of the sub-path from the target ring to an outlet.

Another mismatch between the design and the protocol results from fluid-multiplexing operations. Fluid-multiplexing refers to fluid transportation from the same source to multiple destinations, which is common in high-throughput bio-applications. Though fluid-multiplexing is typically specified as a single operation in current high-level protocols, it may require a dynamic change of the flow path if the flow channels connected to the different destinations are of different lengths.\(^3\) For example, Figure 3(a) shows a photo of a biochip [18] containing five ring-shaped mixers that support fluid-multiplexing from M1 to M2, M3, M4, and M5. When the transportation from M1 to its nearest mixer M3 finishes, the transportation to the other target mixers will still be in progress. In this case, the control channel pressurization protocol needs to be updated to block the fluid motion in M3 without disturbing other sub-paths. Neglecting this feature can lead to inaccuracy of the scheduling results, difficulty in the execution process, and most importantly, incorrect control layer design that fails to support the target application.

Last but not least, due to the absence of channel-level protocols, current physical synthesis methods cannot precisely estimate the usage of control resources, which leads to redundancy in the design. For example, Figure 3(b) shows a design synthesized by a state-of-the-art tool [8], which places a valve at each branch of the flow channels to control fluid transportation in all directions. However, the functionality of the valve at the flow channel branch close to the mixer completely overlaps with the right separation valve of the mixer, and thus can be removed without hurting the performance of the chip. Removing redundant control components including valves, channels, and pins means cleaner control layer design and chip size reduction. Besides, since control channels are much thinner than flow channels, they are more vulnerable to damages and are thus a major source of chip defects. Thus, avoiding redundancy also contributes to the robustness of the chip.

In this work, we propose to bridge the gap between current high-level and physical synthesis methods with a simulation-based approach named VOM. It takes a biochip design and a high-level protocol as inputs, and then synthesizes and optimizes channel-level pressurization protocols to support dynamic construction of valid flow paths. The contribution of VOM includes:

- It is the first approach that synthesizes channel-level protocols, which provides a basis for validating the dynamic construction of flow paths.
- It proposes an event-driven mechanism to automatically update the channel-level protocol to support fluid-multiplexing.
- It optimizes the channel-level protocol based on adjustable criteria including execution time and resource usage.
- It detects the design redundancy and locates the conflicts for incompatible designs and protocols, which opens up a new direction to improve the performance and the feasibility of customized biochip synthesis.

\(^1\)Since PDMS is gas-permeable [15], given large-enough pressure from $i$, the fluids may slowly push the air out of the channel, but the transportation time will be significantly prolonged and the fluids may be mixed with air bubbles.

\(^2\)Because of the area confinement, current physical synthesis methods usually trade off the length-matching feature for routability. Since the application execution time is dominated by biological phenomena but not fluid transportation, this trade-off is considered acceptable [17].
A. An Overview of VOM

In this section, we will briefly introduce the basic building blocks of VOM to provide a quick overview of our algorithmic flow. The system diagram is shown in Figure 4.

1) Inputs

VOM takes a design and a high-level (operation- and device-level) protocol as input.

- The design specifies the physical features of a given multilayered biochip. VOM interprets the flow layer structure into a weighted graph $G = (V, E)$ consisting of vertices $V$ and undirected edges $E$. Each vertex $v \in V$ represents a ‘landmark’ on the chip, which can be an inlet, an outlet, a valve, or a flow channel intersection. Each edge $e \in V$ represents a flow channel between the landmarks, with weight coefficients $w_e$, $h_e$, and $l_e$ representing the dimensions of this flow channel. Besides, the design also specifies the control channel connection among valves.

- The high-level protocol specifies a sequence of fluid transportation operations. For each operation, it describes the target flow path as a set of inlet vertices that initiate the fluid movement, a set of destination vertices at which fluid movement ends, and a set of vertices that should be excluded from the flow path.

2) Flow Layer Compatibility Check

Neglecting the control channel connections, VOM first performs a quick check to determine whether the target flow paths can be supported by the flow layer design. If the check fails, VOM outputs a warning message indicating that the flow-layer design is incompatible with the protocol. Otherwise, VOM proceeds to the next step.

3) Control Sequence Optimization

In this step, VOM restores the channel connections among valves based on the original control-layer design, and looks for all candidate control channel pressurization protocols that can construct the target flow path. If the construction fails, VOM outputs a warning message indicating that the control-layer design is incompatible with the protocol, i.e. some control channel connections are improper. Otherwise, VOM carries out an optimization process based on user-defined criteria to finalize a channel-level protocol from all candidates.

4) Simulation

Based on the graph $G$ and the channel-level protocol, VOM simulates the application execution process in an event-driven manner, which allows it to predict the demand for protocol updates in fluid-multiplexing operations. If such demand is detected, VOM automatically updates the high-level protocol and turns back to synthesize the channel-level protocol that supports the update.

5) Output

If the design and the high-level protocol are compatible, VOM outputs three types of results: a channel-level protocol where the dynamic formation of all flow paths is validated; a schedule for each fluid transportation operation; and a report on resource usage in the control layer.

B. Flow Path Validation

Before diving into the detailed synthesis flow of VOM, we first introduce our flow path validation algorithm, which is an important module frequently applied in most of the synthesis steps.

We perform flow path validation based on a graph $G = (V, E)$ that models the physical structure of a chip, and a set of vertices $V_{\text{block}} \subseteq V$ that specifies the initial blockage in the chip.

As the first step, we assign an availability for each vertex $v \in V$ and each edge $e \in E$ to indicate whether the vertex/edge can be used to form a valid flow path. All edges are initialized as available by default, and for each vertex $v \in V$, if $v \in V_{\text{block}}$, it is initialized as unavailable, otherwise it is initialized as available.

We then propose an algorithm to recursively modify the availability of all vertices and edges to locate the valid flow paths. To guide the modification, we formulate the following rules:

1) If an edge consists of an unavailable vertex, the edge is unavailable.

2) If a vertex belongs to an unavailable edge and the vertex does not represent a flow channel intersection, the vertex is unavailable.

3) If a vertex belongs to only one available edge and the vertex is not an inlet/outlet, the vertex is unavailable.

4) If a vertex is not reachable from the inlet or it cannot reach the outlet, the vertex is unavailable.

Figure 5 demonstrates our modification process with an example, where the design consists of 14 vertices, among which vertices 8 and 12 are specified as initially unavailable, as shown in Figure 5(a).

The first modification rule is based on the fact that the blocked end of a flow channel can neither initiate nor receive a fluid flow. As a result, we specify the edges connected to vertices 8 and 12 as unavailable, as shown in Figure 5(b).

The second modification rule indicates that a chip component without valid flow channel connection is invalid. Thus, as an endpoint of the unavailable edge $(12, 14)$, vertex 14 is specified as unavailable, as shown in Figure 5(c).

The third modification rule follows the fact that an intermediate point in the flow path must be connected to at least two unblocked flow channels. Since vertex 10 only has one valid flow channel connection and it is neither an inlet nor an outlet, it is specified as unavailable, as shown in Figure 5(d).

The fourth modification rule indicates that a valid flow path must include an inlet and an outlet to ensure proper pressure drop. As a
result, vertices 2, 4, 5, and 6 are specified as unavailable, as shown in Figure 5(e).

By recursively applying the modification rules until no more vertex or edge can be specified as unavailable, we can locate a valid flow path, as shown in Figure 5(f).

III. VOM SYNTHESIS FLOW

Based on the system diagram and the flow path validation approach, this section gives a detailed description of each VOM synthesis step.

A. Flow Layer Compatibility Check

The first step of VOM is to quickly check whether a target flow path can be supported by the given flow-layer design.

A target flow path is specified in the input protocol as a set of inlet vertices \( V_{\text{inlet}} \), one or multiple destination vertices \( V_{\text{target}} \), and a set of vertices that should be excluded from the flow path (to avoid contamination) \( V_{\text{exclude}} \).

VOM carries out the check with flow path validation.

1) Specify \( V_{\text{block}} \)

To find out whether fluids can arrive at the destination vertices \( V_{\text{target}} \) without contaminating \( V_{\text{exclude}} \), we initially add all vertices in \( V_{\text{exclude}} \) to \( V_{\text{block}} \) to indicate that these vertices are not available for forming the flow path. Besides, we also add inlet vertices that do not belong to the current flow path to \( V_{\text{block}} \) to prevent them from contamination.

However, not all vertices in \( V_{\text{block}} \) have the risk of contamination. Specifically, a flow path consists of two sub-paths: a sub-path from an inlet to the destination, and a sub-path from the destination to an outlet. Since the fluid transportation will be stopped when fluids arrive at their destination, the latter sub-path will not be contaminated. Adding contamination-free vertices into \( V_{\text{block}} \) leads to two issues:

1) It may hurt the feasibility of a valid flow path. Take the chip shown in Figure 5(a) as an example. For a given protocol \( V_{\text{inlet}} = \{1\} \), \( V_{\text{target}} = \{7\} \) and \( V_{\text{exclude}} = \{8,11,12\} \), which aims to transport fluids from inlet 1 to the flow channel between 3 and 7 while keeping all other channels contamination-free, it is safe to specify \( V_{\text{exclude}} \) as empty since all vertices except for 1, 3 and 7 are in the contamination-free sub-path. However, if we add the whole set of \( V_{\text{exclude}} \) to \( V_{\text{block}} \), there will be no valid flow path left in the design, since vertex 7 cannot reach any outlet.

2) It may prolong the time needed for the transportation. Still using the above example, if we leave vertex 11 available and add 8 and 12 to \( V_{\text{block}} \), though a valid flow path can be constructed as shown in Figure 5(f), the hydraulic resistance of the flow path will be enlarged compared to keeping \( V_{\text{block}} \) empty. Thus, fluids will take more time to reach their destination.

VOM avoids these issues by transforming \( G \) into a directed graph, based on which we can derive whether a vertex is in a contamination-free sub-path. If a vertex in \( V_{\text{block}} \) has no risk of contamination, we remove it from \( V_{\text{block}} \).

2) Check Compatibility

With the design graph \( G \) and the set \( V_{\text{block}} \), VOM performs flow path validation to locate all available vertices and edges. If for all \( v \in V_{\text{inlet}} \cup V_{\text{target}} \), \( v \) remains available after the validation, we can conclude that the flow-layer structure is capable of supporting the target flow path, and VOM proceeds to the next step. Otherwise, VOM outputs a warning message.

B. Control Sequence Optimization

If the input passes the flow-layer compatibility check, VOM first collects all control channel pressurization protocols that can support the target flow paths, and then decides one final protocol based on user-defined optimization criteria.

\(^3\) More detailed discussion about hydraulic resistance will be introduced in Section 3.2.

Figure 5: An example of the recursive modification procedure for locating all valid flow paths. In-/outlets are denoted as circles and valves are denoted as rounded rectangles.
1) Collect Control Channel Pressurization Options

VOM builds a search tree to efficiently traverse all control channel pressurization options. The algorithm is inspired by the idea of branch and cut for solving integer linear programming problems [20].

Except for a virtual root (level-0), each node in the search tree represents a control channel pressurization option, where a level-1 node is a set of cardinality 1 representing an option of pressurizing one control channel, a level-2 node is a set of cardinality 2 representing an option of pressurizing two control channels, etc. Thus, for a design that consists of \( n \) control channels, there will be \( n \) level-1 nodes in the tree.

We then perform a breadth-first search to traverse the tree. For each node, we perform flow path validation with \( V_{\text{block}} \) defined as all valve vertices contained in the control channels of the node, i.e., we assume all control channels in this option to be pressurized and look for the resulting flow paths. Based on the validation results, we assign the node to one of three different classes:

1) The resulting paths do not contain all vertices in \( V_{\text{inlet}} \) and \( V_{\text{target}} \).
   In this case, we classify the node as \( \text{fail} \), which represents an invalid option.

2) The resulting paths contain all vertices in \( V_{\text{inlet}} \) and \( V_{\text{target}} \), and do not contain any vertex in \( V_{\text{exclude}} \). In this case, we classify the node as \( \text{success} \), which represents a valid option.

3) The resulting paths contain all vertices in \( V_{\text{inlet}} \) and \( V_{\text{target}} \), but they also contain some of the vertices in \( V_{\text{exclude}} \). In this case, we classify the node as \( \text{candidate} \).

After traversing all nodes on the same level, for each node that is classified as \( \text{fail} \) or \( \text{success} \), we cut all nodes that represent a superset of this node from the tree. Because if a node is classified as \( \text{fail} \), it means that this option leads to unwanted blockage of the flow paths. Thus, options containing the same set of control channels will also lead to the same unwanted blockage. On the other hand, if a node is classified as \( \text{success} \), it means that this option is enough to support the protocol. Thus, it is also unnecessary to include additional control channels.

After traversing all nodes in the tree, we collect the nodes classified as \( \text{success} \) as potential control channel pressurization options.

Figure 6 shows an illustration of our synthesis method for a design consisting of four control channels. We first build a search tree, and then perform flow path validation for each of the level-1 nodes. Based on the validation results, \( \{c_1\} \) is classified as \( \text{success} \) and \( \{c_2\} \) is classified as \( \text{fail} \). Thus, we cut all the supersets of \( \{c_1\} \) or \( \{c_2\} \) from the tree, which leaves \( \{c_3,c_4\} \) as the only level-2 node. After another flow path validation process, \( \{c_3,c_4\} \) is classified as \( \text{success} \). Thus, we collect it together with \( \{c_1\} \) as the candidates.

2) Optimization Criteria

VOM provides two optimization criteria for selecting a control channel pressurization protocol from all candidates. A user of VOM can choose one of the criteria according to his/her demands.

a) Resource-Oriented Optimization

The first criterion is to execute the application with a minimized number of control channels. Control channels that are never pressurized during the whole application can be considered as redundant and thus removed from the chip. Since the removal of a control channel also results in the removal of all valves it contains and the control pin

![Diagram of search mechanism](image)

Figure 6: Search and collect all valid control channel pressurization options

it is connected to, by minimizing the number of pressurized control channels, we minimize the control-layer redundancy.

We propose an integer-linear-programming method to solve this optimization problem. Specifically, we model the problem as follows:

**Input:**
- A set \( C \) of natural numbers representing the indices of all control channels in a design.
- A sequence \( O = o_1,\ldots,o_n \) representing all fluid transportation operations in the application.
- A function \( f \) that maps each operation \( o \in O \) to a set of control channel pressurization options \( \{C_1,\ldots,C_m\} \) where \( C_1,\ldots,C_m \subseteq C \).

**Output:**
- A set \( M \subseteq C \) representing the control channels that have been pressurized in the application. In other words, \( M \) satisfies:
  \[
  \forall o \in O \exists M' \subseteq M : M' \in f(o)
  \]

**Optimization objective:**
- Minimize the cardinality of \( M \).

To build the model, we introduce a binary variable \( b_{o_j} \) for each \( c_i \) in \( C \) to indicate whether the channel is pressurized in the application. Also, we introduce a binary variable \( b_{o_j,c_k} \) to represent whether a control channel pressurization option \( C_k \subseteq f(o_j) \) is selected to execute operation \( o_j \).

We introduce the following constraint for each \( o_j \in O \) to model that exactly one control channel pressurization option must be chosen for each operation:

\[
\sum_{C_k \subseteq f(o_j)} b_{o_j,c_k} = 1.
\] (1)

We introduce the following constraints to model that if an option \( C_k \subseteq f(o_j) \) is chosen to execute an operation \( o_j \), all control channels
For a rectangular channel of height $h$ and width $w$ in multi-layered continuous-flow microfluidic biochips, its hydraulic resistance is approximated as:

$$R_H = \frac{12\mu L}{h^3 w},$$  \hspace{1cm} (4)

where $\mu$ is the viscosity of the to-be-filled fluids and $L$ is the length of the channels.

Analogous to an electric circuit, for $n$ channels with individual hydraulic resistance $R_1, \ldots, R_n$ that are placed in series, the effective resistance $R_s$ is calculated as:

$$R_s = R_1 + \cdots + R_n,$$  \hspace{1cm} (5)

and if those channels are placed in parallel, the effective resistance $R_p$ is calculated as:

$$R_p = \frac{1}{\frac{1}{R_1} + \cdots + \frac{1}{R_n}}.$$  \hspace{1cm} (6)

For each control channel pressurization option, we perform flow path validation to compute its resulting flow path and calculate its hydraulic resistance. Specifically, we build hyper-edges for edges that are connected in parallel or in series, where the resistance of a hyper-edge is calculated after the resistance of all its edge-segments has been calculated. To note is that a hyper-edge can be built upon other hyper-edges to represent a multi-level fluid-multiplexing structure.
Compatibility check to begin the synthesis of the corresponding channel-level protocol. Whenever a new simulation process begins, the fluid status simulated by the previous process will be inherited and the event-driven protocol update mechanism will be repeated.

### IV. Experimental Results

We implemented VOM in C++, and tested its performance with three multilayered continuous-flow biochip designs.

- The first design consists of a rectangular chamber and a ring-shaped mixer, as shown in Figure 8. Except for four sequentially connected pumping valves, all valves in this design are directly connected to control pins and thus do not share the pressure with the others.

- The second design consists of three ring-shaped mixers, as shown in Figure 9, where valves of similar functionality but in different mixers are sequentially connected to share the same control pins.

- The third design consists of two rectangular chambers and two ring-shaped mixers. It is an application-specific design proposed in [7] and shown in Figure 2(b). It also applies an aggressive pressure-sharing strategy to reduce the number of control pins on the chip, which results in a relatively larger number of sequentially connected valves.

Table I shows the input features and the synthesis results. In the following we analyze the results in detail:

For the first design, we tested five fluid transportation operations from different inlets to the chamber and to the mixer. VOM validates the flow paths for all five operations with channel-level pressurization protocols, based on which it detects two redundant control channels that can be safely removed from the design, as shown in Figure 8.

For the second design, we tested two input protocols:

- The first protocol consists of six fluid transportation operations, each of which starts from an inlet and ends at a half-ring-shaped channel inside a mixer. VOM validates the flow paths for all six operations with channel-level pressurization protocols, and the synthesis results show no redundancy.

- The second protocol consists of two fluid-multiplexing operations, and thus is more in accordance with the intention of the design (since the design consists of three parallel mixers). For each fluid-multiplexing operation, VOM automatically updates the control channel pressurization protocols twice based on the simulation results. Figure 9 illustrates the updates for the first fluid-multiplexing operation, which aims to transport fluids from an inlet to three half-ring-shaped mixer-segments simultaneously. At the beginning of the operation, two control channels are pressurized to form the flow paths without contaminating the other halves of the mixers, as shown in Figure 9(a). When the channel segment in the nearest mixer is filled, VOM detects that there are still two other sub-paths in progress, as shown in Figure 9(b). Thus, it pressurizes another control channel to stop the fluid transportation in the finished sub-path without disturbing the others, as shown in Figure 9(c). Similarly, after fluids reach the middle mixer, VOM pressurizes another control channel to ensure the longest sub-path, as shown in Figure 9(d). Besides, by applying the fluid-multiplexing protocols, three control channels can safely be removed.

The third design is application-specific. Thus, we generated an input protocol consisting of five fluid transportation operations based on the bio-application [24]. The protocol includes a fluid-multiplexing operation, for which VOM updates the channel-level protocol once. When VOM tries to validate the flow path for the third operation from an inlet to one of the mixers, it identifies that there is no control channel pressurization option that can form the required path, due to two sequentially connected valves in two conflicting sub-paths. Thus, we added the fifth test case assuming one additional control channel is added to pressurize the conflicting valves separately. In this case, VOM validates the flow paths for all five operations with channel-level pressurization protocols and detects two redundant control channels.

For all three designs, VOM synthesizes the channel-level protocols within seconds. The program run time increases with the scale of the design and the protocol.

### V. Conclusion

Design automation for multilayered continuous-flow microfluidic biochips has made significant progress in the last decade. However, there is a mismatch between the state-of-the-art high-level synthesis and physical design approaches, which hinders the development of a complete automatic synthesis flow. This work proposes VOM to build this missing link with flow path validation and control sequence optimization. VOM takes a biochip design and a high-level manipulation...
protocol as inputs, and then synthesizes and optimizes channel-level pressurization protocols to validate the dynamic formation of required fluid transportation paths. For fluid-multiplexing operations, VOM automatically updates the high-level protocol, and synthesizes the corresponding channel-level protocol based on an event-driven simulation approach. We demonstrate the efficiency and the effectiveness of VOM with multiple biochip designs.

In general, VOM can efficiently generate channel-level pressurization sequences for feasible designs and protocols, and locate the conflicts for infeasible designs and protocols. It can also detect redundant resource usage of the input design, which opens up a new direction to improve the performance and the feasibility of customized biochips.

REFERENCES


